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### Tissue Processing Presented by: DAYNOVOV ABBOSJON

Research



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### Introduction

- There are 3 main techniques which are used in preparing microscopical sections from tissues:
- The paraffin technique (It is the most **common** method)
- The celloidin technique (It is the most **perfect** method)
- The freezing technique (It is the most **rapid** method)



### ...Introduction

- Tissues from the body taken for diagnosis of disease processes must be processed in the histology laboratory to produce microscopic slides that are viewed under the microscope by pathologists.
- The techniques for processing the tissues, whether
  - Biopsies,
  - Larger specimens removed at surgery, Or
  - Tissues from autopsy are described below.
- The persons who do the tissue processing and make the glass microscopic slides are **HISTOTECHNOLOGISTS**.

### **Specimen Accessioning**

Tissue specimens received in the surgical pathology laboratory have a **request form** that **lists the patient information** and **history** along with a **description of the site of origin.** 

The specimens are accessioned by **giving** them a number that will identify each
 specimen for each patient.





### **Gross examination**

- Tissues removed from the body for diagnosis arrive in the Pathology Department and are examined by a pathologist, pathology assistant, or pathology resident.
- Gross examination consists of describing the specimen and placing all or parts of it into a small plastic cassette which holds the tissue while it is being processed to a paraffin block. Initially, the cassettes are placed into a fixative.





### ...Gross examination

#### • Note:

- When a **malignancy** is suspected, then the specimen is <u>often covered</u> with **ink** in order to mark <u>the margins of the specimen</u>.
- Different colored inks can be used to identify different areas if needed.
- When sections are made and processed, the ink will mark the actual margin on the slide.



### **Tissue Processing steps**

- Biological tissues are generally rather soft, making it quite difficult to cut acceptably thin sections directly from the fresh or fixed tissues.
- Methods must be used to hold the tissues firm, which <u>facilitates</u> cutting thin sections with a sharp knife.
- Firmness can be achieved <u>either</u> by **embedding** the tissues in a suitable **embedment** or by **freezing the tissue**.
- Once the tissue has been fixed, it must be **processed** into a **form** in which it can be made into **thin microscopic sections**.

### ... Tissue Processing steps

• The usual way this is done is with **paraffin**.

- Tissues embedded in paraffin, which is similar in density to tissue, can be sectioned at anywhere from 3 to 10 microns, usually 5-8 routinely.
- The technique of getting fixed tissue into paraffin is called **tissue processing**.
  The main steps in this process are **dehydration** and **clearing**.

### The paraffin Technique

#### • Washing

 Following fixation, the tissues should be washed from 15 to 30 minutes. The fixed tissues are washed in running tap water to remove the fixative from them.

**Dehydration** 

Wet fixed tissues (in aqueous solutions) cannot be directly infiltrated with paraffin.

- **First**, the **water** from the tissues must be <u>removed by dehydration</u>. This is usually done with a series of **alcohols**; say **70% to 95% to 100%**.
- The organic solvent must replace the water **gradually** to <u>prevent</u> **turbulence** at the interface between water and pure ethanol
- Turbulence could cause damage or distortion to cellular components.
  The number of steps or the gradient differences should be determined by
  - The degree of fixation
  - The delicacy of the tissue
  - The degree of cellular detail to be preserved
- Sometimes the first step is a mixture of formalin and alcohol.
- Other dehydrants can be used, but have major disadvantages.
- Acetone is <u>very fast</u>, but a <u>fire hazard</u>, so is safe only for small, hand-processed sets of tissues.
- **Dioxane** can be used <u>without clearing</u>, but has <u>toxic fumes</u>.

#### Clearing

- The next step is called "clearing" and consists of removal of the dehydrant with a substance that will be miscible with the embedding medium (paraffin).
- The commonest clearing agent is **xylene**.
- Toluene works well, and is more tolerant of small amounts of water left in the tissues, but is <u>3 times more expensive</u> than xylene.
- **Chloroform** used to be used, but is a <u>health hazard</u>, and is <u>slow</u>.
- **Methyl salicylate** is rarely used because it is expensive, but it smells nice (it is oil of wintergreen).
- **Excessive** exposure to clearing reagents may cause excessive hardness or shrinkage.

#### <u>Impregnation in paraffin</u>

 The tissues are put from 6 – 24 hours in hot soft paraffin at 50°C, then in hot hard paraffin at 55 °C in the oven. The paraffin will penetrate in-between the cells of the tissues. This process of paraffin infiltration is a necessary step to harden the tissues before their embedding.

- Embedding
- <u>Finally, the tissue is infiltrated with the embedding agent,</u> <u>almost always paraffin.</u>
- **In early days** of microscopy histologists tried to harden tissues artificially with **fixatives**, in order to be able to cut suitably thin sections for microscopy.
- Nearly 100 years ago, the method of embedding tissues in paraffin was developed
- **Paraffin** is a derivative of **crude petroleum**. It is a group of <u>variable</u> <u>length</u>, <u>long-chain</u> **hydrocarbons of the methane** series
- Most paraffins suitable as embedding media melt between 52° and 58°C.
- Since most paraffin have a melting point between 52-58°C, it must infiltrate the cells while it is hot.

- **Infiltration** must be carried out at only a few degrees above the melting point of paraffin
- This represented a **great step forward** in microscopic techniques.
- Firmness was achieved with a supporting medium (an embedment), rather than by hardening the tissue itself.
- For many years paraffin served as almost the only embedment.
- Most of our knowledge from microscopy has been gained from sections cut from paraffin-embedded tissues.

- Paraffin can be purchased that **differ in melting point**, for various hardnesses, depending upon the way the histotechnologist likes them and upon the climate (warm vs. cold).
- **Recently** a product called **Paraplast Plus** was introduced into the market. It contains added **plasticizers** that <u>make the paraffin blocks</u> <u>easier for some technicians to cut.</u>
- A **vacuum** can be applied inside the tissue processor to assist penetration of the embedding agent.

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- The **time required** for embedding tissues using <u>ethanol dehydration</u> and <u>xylene</u> clearing usually **exceeds eight hours**.
- Normally, the tissue is processed overnight with an automatic tissue processing machine.

- Using a completely different rationale for dehydration,
   Prento (1978) was able to reduce the time required for embedding fixed tissues to less than 3 hours, using far few steps.
- He used **Dimethoxypropane** (**DMP**), which served as both the <u>dehydrating</u> and <u>clearing</u> agent.
- Acidified DMP does not simply replace the water but chemically reacts with water to form <u>methanol</u> and <u>acetone</u> which both act as **dehydrants**.
  - After the block of tissue has been completely infiltrated with paraffin, it is placed in a mold containing hot paraffin and oriented in the desire manner.
    The paraffin is then allowed to solidify.

- Upon solidifying, paraffin shrinks 16.5 percent in volume. Paraplast is supposed to shrink less (14 percent by volume)
- No doubt the two most objectionable aspects of paraffin as an embedding medium are:
  - The heat required for melting- the critical shrinkage point of collagen is approximately 65°C. Exposure of collagenous tissues to this temperature must be carefully guarded against to avoid excessive shrinkage.
  - Shrinkage upon solidification
- Despite these problems, paraffin has been far the most widely used embedding medium for many years, and it will probably not be readily replaced by another medium.

- The above processes are almost always automated for the large volumes of routine tissues processed.
- Automation consists of an instrument that moves the tissues around through the various agents on a preset time scale.
- The "**technicon**" tissue processor is one of the commonest and most reliable (a mechanical processor with an electric motor that drives gears and cams), though no longer made.
- Newer processors have computers, not cam wheels, to control them and have sealed reagent wells to which a vacuum and/or heat can be applied.



Tissues that come off the tissue processor are still in the cassettes and must be manually put into the blocks by a technician who must pick the tissues out of the cassette and pour molten paraffin over them. This "embedding" process is very important, because the tissues must be aligned, or oriented, properly in the block of paraffin.





### **Alternatives to paraffin embedding**

- Alternatives to paraffin embedding include various **plastics** that allow thinner sections. Such plastics include:
- Methyl Methacrylate,
- Glycol Methacrylate (GMA),
- Araldite
- Epon.
- Methyl Methacrylate is very hard and therefore good for embedding undecalcified bone.
- **Glycol Methacrylate** has the most widespread use since it is the easiest to work with.
  - **Araldite** is about the same as <u>methacrylate</u>, but requires a more complex embedding process.
    - **Epon** is <u>routinely used</u> for **electron microscopy** where very thin sections are required



#### • Note:

- Plastics <u>require special reagents</u> for dehydration and clearing that are <u>expensive</u>.
- For this reason, and because few tissues are plastic embedded, the processing is usually done by hand.
- A **special microtome** is required for sectioning these blocks.
- Small blocks must be made, so the technique lends itself to small biopsies, such as **bone marrow** or **liver**.

### The freezing Technique

- In this method, the fresh or fixed tissues are frozen hardened and cut with a freezing microtome in the cryostat apparatus within few minutes
- It is a quick and simple method which is commonly used during operations for rapid diagnosis of tumors e.g. carcinoma
- The chemistry of tissues is preserved because we use no heat and no chemical solvents
- Can be used in Histochemistry to demonstrate enzymes and chemical components of tissues
- **Disadvantage**: It gives not-serial thick sections which may fragment into small pieces, so they are very difficult to be stained and to be stored.

**"Floaters**" are <u>small pieces</u> of tissue that appear on a slide that do not belong there--they have floated in during processing.

**Floaters** may arise from sloppy procedure on the **cutting bench-- dirty towels**, **instruments**, or **gloves** can have tissue that is carried over to the next case.

Therefore, it is essential that you **do only one** specimen at a time and clean thoroughly before opening the container of the next case.

- If **reusable cassettes** are <u>employed</u>, you must be aware that tissue may potentially be carried over and appear as "floaters" even several days later, when the cassette is re-used.
  - The problem arises when, during embedding, not all the tissue is removed from the cassette. Then, in the cleaning process, not all of the wax is removed. Then, the next person using the cassette does not pay attention to the fact that there is tissue already in the cassette and puts his specimen in it.
  - The floater that appears on the slide will look well-preserved--it should, because it was processed to paraffin.

Always be sure that **you properly identify the tissue**! This means that you make sure that the patient label on the specimen container matches that of the request slip.

- An **accession number** is given to the specimen. This number must appear with the tissue at all times.
  - You must **never submit** a cassette of tissue **without a label**.

You must **never submit** a cassette of tissue with the **wrong label**.

Mislabelling or unlabelling of tissues is courting disaster

- <u>Sectioning</u>
  <u>Frozen Sections</u>
- <u>Staining</u>
- <u>H & E staining</u>
- <u>Coverslipping</u>
- <u>Decalcification</u>
- Artefacts in Histologic Sections

